

AGE-RAGE synergy influences programmed cell death signaling to promote cancer

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Abstract

Advanced glycation end products (AGEs) are formed as a result of non-enzymatic reaction between the free reducing sugars and proteins, lipids, or nucleic acids. AGEs are predominantly synthesized during chronic hyperglycemic conditions or aging. AGEs interact with their receptor RAGE and activate various sets of genes and proteins of the signal transduction pathway. Accumulation of AGEs and upregulated expression of RAGE is associated with various pathological conditions including diabetes, cardiovascular diseases, neurodegenerative disorders, and cancer. The role of AGE-RAGE signaling has been demonstrated in the progression of various types of cancer and other pathological disorders. The expression of RAGE increases manifold during cancer progression. The activation of AGE-RAGE signaling also perturbs the cellular redox balance and modulates various cell death pathways. The programmed cell death signaling often altered during the progression of malignancies. The cellular reprogramming of AGE-RAGE signaling with cell death machinery during tumorigenesis is interesting to understand the complex signaling mechanism of cancer cells. The present review focus on multiple molecular paradigms relevant to cell death particularly Apoptosis, Autophagy, and Necroptosis that are considerably influenced by the AGE-RAGE signaling in the cancer cells. Furthermore, the review also attempts to shed light on the provenience of AGE-RAGE signaling on oxidative stress and consequences of cell survival mechanism of cancer cells.

Keywords $AGEs \cdot RAGE \cdot AGE \cdot RAGE \text{ signaling } \cdot Cell \text{ death } \cdot Cancer$

Abbreviations

AGE	Advanced glycation end products
Akt	Protein kinase B
DAMPs	Damage-associated molecular patterns
ERK	Extracellular-signal-regulated kinase
HMGB1	High-mobility group box 1
JAK	Janus kinase
MAPK	Mitogen-activated protein kinase
NF-kB	Nuclear factor kappa B
NOX	NADPH oxidase

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Nrf-2	Nuclear factor erythroid 2-related factor 2
PAMPs	Pathogen-associated molecular patterns
PI3K	Phosphoinositide 3-kinases
RAGE	Receptor of advanced glycation endproducts
ROS	Reactive oxygen species
STAT-3	Signal transducer and activator of transcription
	3
TLRs	Toll-like receptors

Introduction

Advanced glycation endproducts (AGEs) are the byproducts of metabolism and harmful substances produced nonenzymatically from glycation reaction between proteins, lipids, or nucleic acid with free reducing sugars [1]. AGEs are formed as the result of the reaction between the electrophilic carbonyl group of reducing sugars with free amino group of different proteins, which resulted in the formation of unstable Schiff's bases that eventually undergo further rearrangements to form stable Amadori products. The stable amadori products undergo further modification (oxidation, dehydration, or polymerization) in the presence of transition metals to form more stable AGEs [2]. The production of AGEs increases manifold under the conditions of chronic hyperglycemia and diabetes due to the availability of free sugars [3]. Since the past decade, multiple AGEs have been identified which includes (but not limited to) GOLD (glyoxal lysine dimer), MOLD (methylglyoxallysine dimer), Nɛ-carboxyethyl-lysine (CEL), pentosidine, and non-cross linking Nɛ- carboxymethyl-lysine (CML). Despite having different types or status of cross-linkings, the consequences of all AGEs remain similar in the progression of different pathologies [2].

Although the cross-linked AGEs appear to be inert per se, they exert their effects in the development or progression of different pathologies through their interaction with non-specific surface receptors known as the Receptor of advanced glycation endproducts (RAGE) [4-6]. RAGE is a multi-ligand-specific receptor that binds to AGEs and other danger signaling molecules known as DAMPs to exert their pathophysiological roles in multiple disorders. These DAMPs include High-mobility group box proteins (HMGB1), calgranulins (S100 proteins), and amphoterin, to name a few [6]. RAGE can also activates an innate immune response against microbial pathogen-associated molecular pattern molecules (PAMPs) including bacterial endotoxin, microbial DNA, and viruses. RAGE is expressed at low levels under normal physiology conditions, but it is upregulated under pathological conditions including, diabetes, cancer, and chronic inflammation. The synergy of RAGE with their ligands can influence NF-kB activation and inflammatory responses [7]. The consistent perpetuation of NF- κ B activation and inflammatory responses mediated through the RAGE signaling accelerates oxidative stress and pathological consequences. Moreover, RAGE triggers various signaling molecules that activate downstream signal transduction pathways directed to NF-kB activation and stimulates the production of various cytokines and growth factors that may be responsible for chronic inflammation and progression of cancer.

Synergy of AGE-RAGE signaling and cancer

Cancer is a multifactorial disease, which is associated with multiple aberrant signaling pathways in a non-transformed cell [8]. Due to the aberrant signaling pathways, non-transformed cells acquire various characteristics including limitless replicative potential, genomic instability, evasion to apoptosis, angiogenesis, invasion, and metastasis to become cancerous, which are commonly known as the hallmarks of cancer [9]. Inflammation can contribute to tumorigenicity and tumor promotion by supplying bioactive molecules to the tumor microenvironment, including growth factors that sustain proliferative signaling, survival factors that limit cell death, invasion, pro-angiogenic factors, metastasis, extracellular matrix-modifying enzymes that facilitate angiogenesis, and inductive signals that lead to activation of epithelial-tomesenchymal transition [10, 11]. Furthermore, inflammatory cells can release chemicals, especially reactive oxygen species, which are actively mutagenic for the neighboring cancer cells, accelerating their genetic evolution towards states of heightened malignancy [12, 13]. Therefore, inflammation has been considered as an emerging hallmarks of cancer [14]. Reinforced of cellular reprogramming, activated immune cells orchestrate production of pro-inflammatory cytokines and various signal transduction pathways to continuous cell proliferation of cancer cells [15]. The aerobic glycolysis is one of the major phenotypes of cancer cells that allows cancer cells to adapt higher glucose uptake and thus continuously exposed to hyperglycemic conditions which are majorly involved in the synthesis of AGEs [16]. Apart from its constitutive expression in immune cells and lung tissues, RAGE is inducibly expressed in cancer cells [17]. Binding of AGEs with RAGE activates various signaling pathways including MAPK, ERK1/2, PI3K, Akt, JAK-STAT, and NF-kB associated with cell survival, inflammation, and cancer progression [18]. Besides promoting cell survival and inflammation around the tumor microenvironment, RAGE and its ligands also promote angiogenesis, cell migration, cell proliferation, invasion, and metastasis by restricting cell death from apoptosis [19-21]. Due to insufficient availability of oxygen in the tumor, the cancer cells remain under continuous deprivation of oxygen, which is known as hypoxia. The hypoxic condition promotes angiogenesis, invasion, metastasis, drug resistance phenotype, and restricts apoptotic machinery during cancer progression. Hypoxia also induces AGE-RAGE-mediated activation of HIF1 α , NF-KB, ERK, and Akt signaling pathways which contribute to the progression of cancer [22]. The tumor hypoxia also accumulates RAGE expression. In addition, the HIF1 α (a master regulator of hypoxia) and RAGE are closely associated with cancer. A recent report suggests that the NF-KB-RAGE-KRAS-HIF-1a pathway underlies the progression of pancreatic cancer [23]. Moreover, the involvement of the AGE-RAGE axis has been shown to promote the autophagic flux with simultaneous inhibition of apoptotic signaling in cancer cells. The activation of autophagic proteins such as Beclin-1 promote the survival of cancer cells through autophagy [17]. Activation of AGE-RAGE signaling also generates oxygen-free radicals, causes oxidative stress, and activates NF-kB that secrets pro-inflammatory cytokines, growth factors, and adhesion molecules such as ICAM-1 and VCAM-1 eventually leading to cancer progression. AGEs may alter the extracellular matrix (ECM) through engagement of cell surface receptors and pro-inflammatory

cytokines production, and ROS may lead to oxidative stress and cancer. Synergy of AGE ligand with their receptor is associated with upregulation of vascular endothelial growth factor (VEGF) and metalloproteinase-2 (MMP-2) as well as the disruption of VE-cadherin/catenin complex that may favor angiogenesis [24]. Current report revealed that overexpression of RAGE augments cell migration, invasion, and epithelial-to-mesenchymal transformation in human lung adenocarcinoma cells through ERK signaling [25]. A recent report suggests that the AGEs also promote cell proliferation and cell migration in breast cancer cells [26]. The previous report revealed that activated Akt, PCNA, and MMP signals further support the inhibition of apoptosis in cancer cells [27]. Inhibition of RAGE signaling in cancer cells has been successful in curbing cancer growth in multiple studies [28-30]. The association of AGE-RAGE signaling with various molecular mechanisms involved in the progression of cancer is shown in Fig. 1.

AGEs, RAGE, and ROS

Accumulation of AGEs accelerates the production of reactive oxygen species (ROS) and oxidative burden in cells and tissues. ROS is an array of reactive molecules that play an important role in maintaining cellular homeostasis. ROS regulates various cell signaling pathways directly or indirectly, which regulates various physiological processes including cell proliferation, differentiation, cell death, inflammation, and immunity [31]. ROS is mainly generated either by mitochondrial ETC or through intracellular enzyme systems such as NOX, LOX, Xanthine oxidase, Cyclooxygenase, NOS, and Cytochrome p450 monooxygenase [32]. The cellular ROS generation is also induced by various growth factors, intra and extracellular toxin substance, AGEs, and cytokines. The H_2O_2 and O_2^- are major reactive oxygen species involved in cellular signaling. A low-level of ROS regulates various physiological functions, but an elevated level of ROS leads to oxidative stress and cell damage. The ROS is generally metabolized by intracellular antioxidant enzymes and prevent the adverse effects of excessive ROS in a normal cell. Moreover, ROS target various cellular redox-sensitive proteins and other biological macromolecules and alter their normal functions thereby resulting in the progression of pathological consequences. An elevated level of ROS is associated with several pathologies including cardiovascular diseases, diabetes, neurodegenerative diseases, inflammation-associated injuries, aging, and cancer [33]. Previous reports revealed that the cancer cells accumulate ROS due to high metabolic rate, activity of various enzyme systems, and mitochondrial dysfunction [12]. Moreover, elevated ROS results in oxidative stress and altered gene regulation leading to the progression of abnormal cell growth, proliferation, invasion, and metastasis. The cancer cells maintain a relatively higher redox state compared to normal cells that sustain proliferative pathways and inhibit programmed cell death signaling. Indeed, AGEs-RAGE signaling generates ROS and initiates various signaling pathways during the onset of cancer [34]. Oxidative stress plays a crucial role in the activation of AGEs-RAGE signaling and pathogenesis of

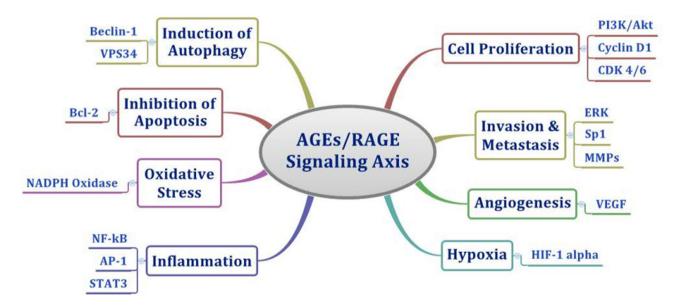


Fig. 1 The association of the AGE-RAGE signaling with molecular mechanisms involved in the progression of cancer. The figure shows an association of the AGE-RAGE signaling with different pathological conditions involved in the progression of cancer including cell

proliferation, invasion, metastasis, and angiogenesis. This even influences cellular redox homeostasis, tumor-associated inflammation, and programmed cell death signaling

diabetes, chronic inflammation, and cancer. Initially, it regulates the downstream pathways and later on it promotes the synthesis of more AGEs [35, 36]. AGEs-RAGE-generated ROS accomplishes cell survival and evades programmed cell death signaling during the progression of cancer. Although, AGEs-RAGE-mediated ROS promotes apoptotic cell death in normal cells, accumulation of AGEs and RAGE promotes NF-κB activation, release of pro-inflammatory cytokines, and oxidative stress that may cause various pathological consequences. Accumulation and activation of AGE is one of the key pro-inflammatory factors in the progression of cancer, diabetes, and diabetic retinopathy. Synergy of AGE-RAGE induces the activation of MAPK signal transduction pathway with NOX-mediated ROS generation and activation of NF- κ B, which leads to upregulation of transcriptional activity of target genes including growth factors (VEGF), adhesion molecules (ICAM-1, VCAM-1), and pro-inflammatory cytokines (IL-6, IL-1 β , TNF- α) and chemokines MCP-1) [37]. Role of ROS in AGEs-RAGE signaling is an emerging interest to shed light on insight molecular mechanisms to understand the complex biology of programmed cell death and survival signaling in cancer.

Synergy of AGE-RAGE influences programmed cell death signaling in cancer

Programmed cell death (PCD) is a physiologically conserved process for the development and removal of unwanted damaged cell for maintaining cellular homeostasis. Programmed cell death is broadly categorized into three types: Type I-Apoptosis, Type II-Autophagy, and Type III-programmed necrosis. Different types of cell deaths exhibit multiple cellular phenotypes that affect many intracellular organelles, cell nucleus, and membrane [38]. Apoptotic cell death (Type I) eliminates the damaged, harmful, unwanted, and senescent cells and is also involved in the function and regulation of the immune system, differentiation, homeostasis/proliferation, and development. Apoptotic cell death is executed by two major pathways, namely, extrinsic and intrinsic apoptotic pathways via caspase cascades to drag the cells towards death [39]. Autophagic cell death (Type II) is the catabolic process that acts as a quality control mechanism for organelles and proteins to favor survival during starvation or scarcity conditions [40]. Necroptosis or programmed necrosis (Type III), is a caspase-independent cell death that is stimulated by tumor necrosis factor receptor 1 (TNFR1), TLRs, IFN receptors (IFNR), and intracellular RNA- or DNA-sensing molecule upon impairment of apoptosis [41]. Necroptosis induces an innate immune response against the infection caused by viruses [42]. All these orchestrated mechanisms of cell death get altered under pathophysiological condition such as cancer. The deficiency of cell death is significantly involved in tumor development and resistance against radiations as well as chemotherapy [38]. ROS plays an intermediate role in cell death and survival signaling. An intracellular level of ROS determines the fate of the cell. Therefore, induction of cell death by manipulating the redox balance in the cancer cells has been considered as one of the promising approaches for cancer therapy.

The AGE-RAGE signaling and apoptosis

The role of AGEs and its receptor RAGE has been investigated for induction of apoptosis in different types of cells [43–47]. The propagation of apoptosis through the AGEs-RAGE axis involves pro-apoptotic factors and caspase cascades. The apoptotic signaling channelizes through death receptor activation and mitochondria disintegration followed by the activation of executioner caspase-3 [48]. The expression of RAGE modulates the death receptor and mitochondrial pathways of apoptosis by regulating the expression of pro-apoptotic caspase-3, caspase-9, and antiapoptotic Bcl-2 [49, 50]. It has been shown that RAGEdeficient cardiac cells have enhanced Bcl-xL expression and reduced cytochrome c release [51]. An earlier investigation also demonstrates that exogenous α -Fas together with AGE aggravate the release of cytochrome c, activation of caspase-8, and caspase-3. Moreover, AGEs may activate Fas-FasL signaling in human retinal ARPE-19 cells [52]. In addition, AGEs-RAGE interaction activates NF- κ B to stimulate TNF- α secretion in cell lines including the macrophages [53] and microvascular endothelial cells [54]. Furthermore, AGE-mediated apoptotic signaling in pericytes orchestrates through initiator caspase-10 and is executed through caspases-3, -6 -7, or 9 [55]. One of the primary mediators through which the AGE-RAGE axis induces apoptosis is elevated reactive oxygen species (ROS) level in the cells [46, 56]. Earlier studies highlighted that AGEs-mediated cell death may influence cellular alterations that may cause pathological complications [57, 58]. The oxidative burden within the cells further perturbs several signaling pathways involved in cellular homeostasis. For instance, AGEs-induced oxidative stress leads to the activation of NF- κ B and MAP kinase (MAPKs) pathways [59, 60]. The MAPKs are a family of serine/threonine kinases that participate in apoptotic signaling. In addition, the JNK and p38 MAPK are the well-versed intermediates that activate apoptosis in response to cellular stress and chemotherapeutic drugs [61-64]. It has been shown that AGEs activate apoptosis via JNK and p38 MAPK in osteoblast [65, 66] and in Schwann and mesangial cells [67, 68]. Furthermore, the AGEs-ROS mediated p38 MAPK and JNK activation and in turn orchestrate pro-apoptotic caspase-3 cascade [60]. Importantly, AGEs have been reported as the pro-apoptotic factors for the cellular culture of microvascular cells, neuronal cells, fibroblasts, and renal mesangial cells [68-71]. In contrast, the AGEs cognate partner S100P/RAGE itself activates apoptosis signaling through the MAPK pathways [72]. On the other hand, RAGE ligand HMGB1 under reduced condition induces Beclin-1-dependent autophagy, while in oxidized state promotes apoptosis [73, 74]. Indeed, ROS triggers cell death signaling through the mitochondrial and endoplasmic reticulum stress-mediated pathway [75, 76]. AGEs-RAGE axis further extends the signaling to mitochondrial functioning. It has been shown that AGE-RAGE interaction leads to mitochondrial dysfunction and altered mitochondrial dynamics [77-79]. The fractions of RAGE have been identified in the extract of mitochondrial complex I and II [80]. The damaged mitochondria accumulate ROS that leads to the disintegration of mitochondrial membrane potential (MMP) and prefers apoptotic instigation in the absence of autophagy signaling [74, 81]. It is observed that AGEs lead to the exacerbation of mitochondria associated pro-apoptotic protein Bax that leads to the mitochondrial alteration and caspase-9 activation in mesangial cells [82]. Interestingly, AGEs-induced disintegration of MMP may be restored with supplementation of glutathione together with an antioxidant N-Acetyl cysteine (NAC) [83].

On the contrary to the pro-apoptotic roles of RAGE in normal cells, cancer cells tend to employ RAGE signals for their proliferative purposes. Apoptosis is the most prominent form of programmed cell death that cancer cells manage to evade by disrupting numerous cellular mechanisms. RAGE and its ligand HMGB1 is known to be upregulated in many types of tumor and promotes invasion and metastasis by activating diverse cellular signaling pathways [84]. The induced expression of RAGE associated with cancer serves multiple purposes to assure the survival of transformed cells via the activation of NF- κ B [2]. RAGE-mediated activation of NF- κ B is associated with angiogenesis, cell migration, proliferation, and evasion of cell death [85]. Multiple studies suggest that RAGE meticulously perpetuates the survival signaling in cancer cells by promoting the "programmed cell survival" (Autophagy) and suppressing "programmed cell death" (Apoptosis) under metabolic and oxidative stress [86]. In a study carried out by Kang R et al. [86], RAGE knockdown resulted in increased apoptosis with diminished autophagy in pancreatic cancer cells under high oxidative stress. Supportively, RAGE overexpression was positively associated with reduced apoptosis and sustained autophagy. Moreover, RAGE overexpression restricted the mitochondrial translocation of p53 and prevented apoptosis [17]. The release of HMGB1 from autophagic cells and its subsequent binding to RAGE support the cancer cell proliferation via ERK1/2 and MAPK activation [87]. HMGB1/RAGE interaction also prevents apoptosis in cancer cells through the activation of Akt and MMP-9 [88]. A study conducted by

Elangovan et al. suggests that downregulation of RAGE by RNA interference (RNAi) reduced the survival of prostate cancer cells by downregulating RAGE as well as its physiological ligand HMGB1 [89]. Our previous report demonstrates that quercetin promotes apoptosis by attenuating the expression of RAGE and its ligand HMGB1 in human breast adenocarcinoma cells [85]. In addition, the quercetin inhibits RAGE/PI3K/Akt/mTOR axis and increases gemcitabine chemosensitivity in pancreatic cancer cells [90]. The sustained autophagic response in cancer cells under stress (along with the inhibition of apoptosis) is also largely attributed to the decreased phosphorylation of mammalian target of rapamycin (mTOR) accompanied by the simultaneous interaction of autophagic protein Beclin-1 with Vps34 [91]. Thus, upregulated expression of RAGE promotes cancer cell survival by sustaining autophagy and inhibiting apoptosis.

A possible explanation for the functional disparity of RAGE in normal and cancer cells to regulate apoptosis can be attributed to the ability of cancer cells to display high intracellular glyoxalase activity. In the case of normal cells, the induced apoptosis is the result of intracellular accumulation of glycation products. However, the survival pathways mediated through the AGE/RAGE axis in cancer cells are mainly sustained through receptor-governed signaling present on the cell surfaces. Moreover, since a high glyoxalase activity is maintained in cancer cells, apoptosis due to the intracellular glycation is seldom observed in such cells. Furthermore, the generation of ROS through AGE/RAGE interaction in normal cells generally results in apoptotic cell death due to free radical-induced DNA damage. However, cancer cells have a characteristic feature to survive under high oxidative stress. Cancer cells scrupulously maintain a relatively higher oxidative burden to regulate their signaling pathways. In that view, AGE/RAGE-mediated oxidative stress can result in apoptosis in normal cells while showing an opposite (anti-apoptotic) effect in cancer cells. (Fig. 2) Taken together, AGE/RAGE-mediated signaling has a close connection in the regulation of programmed cell death.

The AGE-RAGE signaling and autophagy

Autophagic cell death is pertinently suggested as a double-edged sword for programmed cell death and survival. Autophagy is an essential catabolic process for the degradation of cellular components to sustain cellular metabolism under nutritional deprivation conditions. The autophagic process involves various upstream regulators, initiation, and nucleation of autophagophore and autophagolysosome formation, a sequence of events that ultimately aims at the degradation of intracellular components. It plays a vital role in cellular energy balance and homeostasis. However, the defects in autophagic machinery cause several pathologies

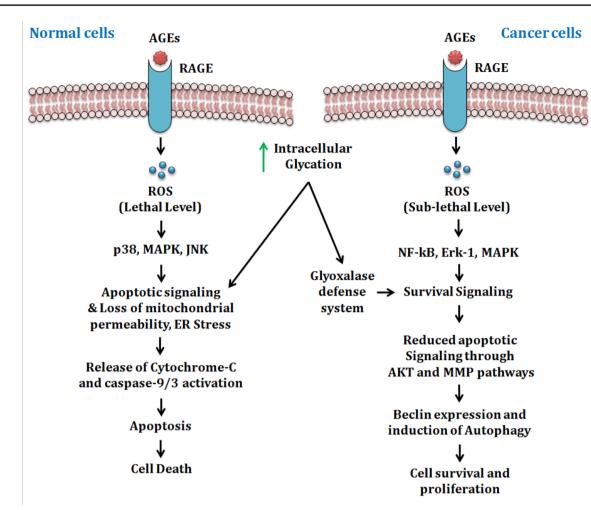


Fig. 2 Role of AGE-RAGE signaling in apoptotic cell death in normal cells and cancer cells. The AGE-RAGE signaling promotes cellular ROS generation and plays an important role in apoptotic cell death in normal cells and cancer cells. Once the AGEs are synthesized it circulates through the system and binds to RAGE. The binding of AGEs to the RAGE activates the number of signaling molecules which promote ER stress and loss of mitochondrial permeability sub-

sequently induces apoptotic cell death in normal cells. On the other hand, the AGE-RAGE signaling promotes survival signaling by inhibition of apoptosis and induction of autophagy in cancer cells. AGE-RAGE signaling may intricate dual role in apoptotic cell death and cell survival signaling. Dysregulated activation of AGE/RAGE signaling may contribute various pathological conditions

including cancer. Moreover, Autophagy has been reported to play dual roles in cancer. Cancer cells possess a high proliferation and growth rate resulting in nutrient stress conditions in the tumor microenvironment. In addition, oncogenic mutations like KRas lead to metabolic adaptation in cancer cells to sustain in the tumor microenvironment [92]. The role of Ras in autophagy in cancer cells largely appears to be paradoxical [93]. A plethora of literature suggests that autophagic signaling acts as an adaptive stress response that enables cancer cells to survive under nutrient-deprived conditions [94]. Additionally, the tumors are deprived of oxygen, forcing them to evolve mechanisms to survive during hypoxic conditions. Hypoxia plays a key role in cancer metastasis and is also intricately related to autophagy machinery in cancer cells [95]. Moreover, autophagy sustains cancer cells under therapeutic stress and enables drug resistance mechanisms. A recent report suggests that the inhibition of autophagy restrains the chemosensitivity in cancer cells [96]. In addition, the induction of autophagy also inhibits apoptosis in cancer cells [97].

AGEs and RAGE are reported to accumulate during the progression of cancer. AGEs participate in autophagy signaling in a wide variety of cells and are involved in cellular dysfunction [98, 99]. Previous reports have highlighted that AGE-RAGE signaling mediates ROS generation and is associated with autophagy in cancer cells to resist apoptosis [22, 100]. In combination, ROS and oxidative stress help to progress autophagy that may favor cancer cells for cell survival and growth [101]. AGEs induce autophagy via the ERK pathway and increase the proliferation of VSMCs. The

knockdown of RAGE in the tumor cell diminishes autophagy and tumor cell survival, which in turn results in the induction of apoptotic cell death. On the contrary, overexpression of RAGE promotes autophagy and diminishes apoptosis to support tumorigenesis. Moreover, RAGE-mediated autophagy is also associated with decreased phosphorylation of mTOR and increased Beclin-1/VPS34 autophagosome formation [17]. In addition, the AGEs-RAGE potently induces autophagy more than the known inducers such as TNF or doxorubicin. However, the potency of AGE-mediated autophagy also depends upon the expression of RAGE. AGE-RAGE activates PI3K, a kinase that plays vital roles in autophagy. Few evidence also suggests that the accumulation of AGEs leads to the ROS generation. NF-kB is intricately involved in autophagy as it controls the transcription of many autophagic genes including Beclin-1. AGE-RAGE signaling also activates PKC and/or RAF kinase and downstream p38/MAPK and ERK pathways to mediate autophagy [102]. As oxidative stress generates oxidative injury and cell death, there are evidences that AGEs activate Nrf-2, a master regulator of antioxidant genes and provide defense against oxidative stress in diabetes [103]. Recent report suggests that the AGEs regulate Nrf-2 and Bcl-xL signaling and promotes survival of oral cancer cells in diabetic patients [104]. It has been also suggested that the AGEs/RAGE signaling axis leads to excessive autophagy and impairs the cell viability of cardiomyocytes, causing cardiac dysfunction [105]. Apart from that, AGEs are also known to mediate autophagy in cardiomyocytes via ERK1/2, whereas prolonged exposure of AGEs to cardiomyocytes leads to apoptotic cell death via the p38/MAPK pathway [106]. Moreover, PI3K/Akt/ mTOR signaling pathway negatively regulates autophagy. AGEs-RAGE interaction also inhibits the PI3K/Akt/mTOR signaling pathway and results in autophagy [107].

Interestingly, RAGE avidly regulates p65 and BNIP3 to promote autophagy and cell survival [105]. A contradictory result is also evident from a study where RAGE-deficient HCC represents the onset of autophagy through the AMPK/ mTOR signaling pathway [108]. The mechanistic pathways that confer to autophagy via AGE-RAGE interactions also show large diversity and remain to elucidate. The Role of AGE-RAGE signaling in pro-survival autophagy during cancer progression is shown in Fig. 3.

The AGE-RAGE signaling and necroptosis

Necroptosis is a caspase-independent cell death that is morphologically analogous to necrosis, but mechanically resembles to apoptosis [109]. Oxidative metabolism generates reactive oxygen species (ROS) via mitochondrial respiratory chain and redox-active during the execution of cell death via necroptosis. AGEs are formed due to the synthesis

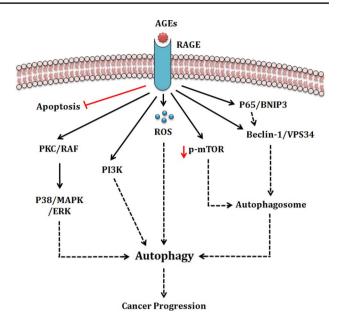


Fig. 3 Role of AGE-RAGE signaling in Pro-survival autophagy during cancer progression. The AGE-RAGE signaling promotes autophagy that mediates cancer cell survival. The binding of AGEs with RAGE activates PKC/RAF, PI3K, and p65/BNIP3 which promotes pro-survival autophagy and inhibits apoptotic signaling in cancer cells. AGE- and RAGE-mediated generation of cellular ROS may contribute stress-associated pro-survival autophagy in cancer

of methylglyoxal, toxic derivatives, dihydroxyacetone phosphate (DHAP), and from the fragmentation of glyceraldehyde-3-phosphate during the process of glycolysis. An interesting study shown that the increased level of cellular glucose possibly induces necroptosis [110]. Receptor-Interacting Protein 1 (RIP1), RIP3, and Mixed Lineage Kinase domain-like (MLKL) are the key mediators of Necroptosis. The assembly of RIP1 and RIP3 result in the formation of the "Necrosome", which is one of the significant characteristics of necroptosis [111]. The subsequent phosphorylation of MLKL by necrosome releases cell damage-associated molecular patterns (DAMPs) with the loss of plasma membrane integrity. The release of DAMPs evokes the inflammatory responses and favors the progression as well as the survival of tumor cells. High-mobility group box 1 (HMGB1) is a distinctive DAMP molecule. The intracellular HMGB1 is present within all nucleated cells and plays a significant role in the nuclear and cellular homeostasis, whereas the extracellular HMGB1 commence and protract the inflammatory response via the ligation of the RAGE and TLRs [112]. Furthermore, the release of HMGB1 and AGEs during necroptosis may interact with RAGE present on the surrounding tumor cells that support inflammation and cancer progression. (Fig. 4) The HMGB1 is a part of the family of the High-mobility group of non-histone chromosomal proteins. It is expressed as a single chain of the polypeptide consisting of 215 amino acids [113]. HMGB1 maintains the

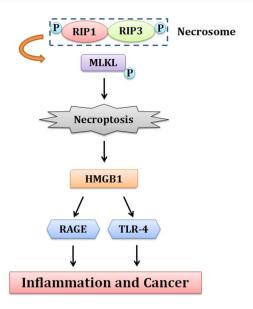


Fig.4 The association of the RAGE signaling with necroptosis during the progression of inflammation and cancer. The intracellular HMGB1 plays a significant role in cellular homeostasis. Upon necrotic insult, the assembly of RIP1 and RIP3 forms necrosome which subsequently releases HMGB1. This extracellular HMGB1 binds with RAGE or TLR-4 that may influence inflammation and cancer progression

chromosomal structure and stability inside the nucleus of a cell. During stress condition, HMGB1 translocates to the cytoplasm from the nucleus to protect the cell from death via mechanism of autophagy by dissociation of Beclin-1/ Bcl-2 interaction for competitive binding with Beclin-1. Once HMGB1 is secreted out or released from the cells, it activates a different set of pathways by binding with Tolllike receptors (TLR-2,TLR-4 and TLR-9) and Receptor of Advanced Glycation Endproducts (RAGE) for induction of inflammation and cell survival [114]. Several cytokines and inflammatory mediators promote tumor growth through TLR-mediated signaling pathways, which lead to activation of transcription factors: NF-kB and STAT-3 for marinating tumor microenvironment consisting of an inflammation [115]. HMGB1 released from necrotic cells orchestrate TLRs and RAGE signaling amplifies inflammatory response by creating functional tripod to maintain chronic inflammatory state [116]. TLRs are evolutionary conserved transmembrane protein found in the cell surface of immune cells. TLRs are key components of the innate immune system that direct detection of pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs) [117]. DAMPs released from injured or dying cells behave as endogenous ligands for TLRs and are recognized by specific TLRs on immune cells; subsequently, TLR signaling will lead towards the inflammatory response. The immune cells including macrophages and dendritic cells consist TLR and RAGE receptor on their cell surface and binding of HMGB1 resulted release of inflammatory mediators including IL-6 and TNF- α . Inconsistent release of IL-6 and TNF- α leads to necrotic or apoptotic cells and maintain the HMGB1 release, resulting a cascade amplification of inflammation [118]. HMGB1 also interacts with TLR5 and initiates MyD88-dependent activation of NF-kB signaling pathway resulting in production of pro-inflammatory and perception of pain [119]. Previous report suggests that the core component of TLR signaling is activation of an IL-1-like pathway dependent upon the adapter MyD88, coupling via an intricate series of kinases and scaffolding proteins to activation of NF- κ B [120].

HMGB1 is a damage-associated molecular pattern (DAMP), which contributes progression of inflammation and cell survival of cancer [121]. HMGB1 is passively released from the damaged cells during necrotic cell death. An active release of HMGB1 during the angiogenic or inflammatory condition promotes cell survival [113, 122]. HMGB1 can activate diverse sets of signaling components, including AKT and MAPKs and play an important role in cell proliferation via synergy of RAGE and hasten cell cycle progression [84]. Interaction of HMGB1 with their receptor RAGE and TLRs influences three major signal transduction pathways including NF-kB, PI3K/Akt, and MAPK/ERK1/2/ p38 [114]. It has been noticed that HMGB1 influences cell proliferation of diffused large B cell lymphoma cells by activation of ERK1/2, Akt, and STAT-3, Non-Receptor Tyrosine Kinase (Src), and SRC Proto-Oncogene. However, obstruction of HMGB1 signaling may inhibits tumorigenesis [123]. Over expression of HMGB1 directly or indirectly influences hallmarks of cancer, that includes the limitless potential to divide, angiogenesis, evasion of apoptosis, compromised sensitivity to growth inhibitors, increased response to growth signals (self-sufficiency), inflammation, invasion, and metastasis [122]. Taken together, release of HMGB1 and other pro-inflamatory cytokines during necrotic insult orchestrates cell survival and enhances the stemness of resident cancer cells for continuous cell proliferation.

Role of miRNAs in regulation of oncogenic AGE-RAGE axis

MicroRNAs (miRNAs) are endogenous, single-stranded family of non-coding RNAs, implicated in regulation of target genes at the transcriptional and translational level [124, 125]. Over the past decade, several studies highlighted the importance of miRNAs in regulation of various human diseases, including, cancer [126–128]. Dysregulated miRNAs signaling in cancer cells up and/or downregulate oncogenes, which, in turn, leads to tumor proliferation, angiogenesis, and apoptotic evasion [129, 130]. Moreover, a systematic profiling of cancer-associated miRNAs provides an informative insight to monitor cancer progression and therapeutic response in patients [131, 132]. The intriguing role of miR-NAs in modulation of cancer gene expression attributed a significant attention in recent past years; however, a mechanistic linkage between miRNAs and pathways associated with tumor progressing remains enigmatic.

Given the importance of miRNAs in cancer gene regulation and AGE-RAGE signaling in cancer proliferation, several studies have identified novel miRNAs involved in the tuning of the AGE-RAGE axis. The oncogenic miR-21, which is reportedly upregulated in various cancer cells, activates though SP100P/RAGE signaling pathway and leads to cancer progression [133, 134]. Indeed, miR-221 and miR-222 is reportedly involved in cell proliferation, through inhibiting cell cycle regulator p27kip1 [135]. In this context, an interesting study showed that HMGB1-RAGE axis induces the oncogenic expression of miR-221 and miR-222 in papillary cancer cell line, to promote cellular malignancy [136]. Moreover, exogenous HMGB1 interacts with RAGE and activates miR-221/miR-222 expression, which in turn, inactivates oncosuppressor PTEN (Phosphatase and tensin homolog) in thyroid carcinoma cell lines, suggesting that HMGB1/RAGE pathway promotes oncogenic miRNAs expression to favor cancer proliferation and targeting these pathways could be a promising approach against tumor growth [137]. An interesting study showed that the treatment of human monocytes with AGEs induces the expression of miR-214, which specifically target tumor suppressor PTEN to delay apoptosis of monocytes [138]. In contrast, an interesting study demonstrated that AGE-induced miR-223 expression target insulin-like growth factor-1 receptor (IGF-1R) expression and leading to apoptosis in osteoblast-like MC3T3-E1 cells [139]. In addition, a dysregulated expression of miR-205 has been inversely associated with the invasive traits of triple-negative breast cancer (TNBC); moreover, ectopic expression of miR-205 in MDA-MB-231 cells significantly target HMGB1/RAGE signaling pathway to attenuate cell growth and metastatic invasion, following studies that suggest that apart from oncogenic transformation, certain miRNAs contributes in regulation of cell proliferation through distinct signaling [140].

RAGE inhibitors as a therapeutic agent

The synergy of AGE-RAGE accomplishes downstream signaling of NF-kB activation and other molecules, which leads to oxidative stress and pathological consequences. Multiple evidences suggested that the elevated expression of RAGE considerably evades cell death in cancer cells. Moreover, the activation of RAGE in cancer cells stimulates diverse sets of signaling pathways that support tumor growth. In addition, AGE-RAGE signaling promotes survival mechanism (prosurvival autophagy) and evades cell death (apoptosis) which appears to be major pathways during cancer progression. In the recent past, several reports highlighted that RAGE inhibitors significantly inhibited tumor growth and induces cell death. An inhibition of AGE-RAGE signaling by natural or synthetic inhibitors can be considered as therapeutic targets. We have explored various databases to find out RAGE inhibitors those are identified to inhibit cell proliferation in

Table 1 RAGE inhibitors that induce apoptosis or inhibit cell survival in various malignancies

RAGE inhibitor	Mechanism involved	Type of cancer	References
Papaverine	Inhibition of RAGE dependent NF-ĸB activation	Human fibrosarcoma	[141]
Curcumin	Inhibition of ERK1/2 and NF-κB, Increases ROS to induce apoptosis	Lung cancer, Nasopharyngeal carcinoma	[2]
Quercetin	Inhibition of HMGB1 and promotes apopto- sis, Increases ROS to induce cell death	Breast cancer	[85, 142]
Withaferin A	Increases ROS to induce cell death	Breast cancer, Head and neck cancer	[2, 143, 144]
Small RAGE antagonistic peptide	Decreases interactions of RAGE ligands (S100s, HMGB1) with RAGE, Inhibition of NF-κB	Pancreatic ductal adenocarcinoma	[145]
Hispidin	Inhibition of NF-κB and RAGE expression	Pheochromocytoma	[146]
Ergothioneine	Inhibition of NF-KB and RAGE expression	Pheochromocytoma	[146]
Ethyl pyruvate	Inhibition of NF-κB, STAT-3, HMGB1, and RAGE expression	Human mesothelioma	[147]
Acetylated apurinic apyrimidinic endo- nuclease 1/redox factor-1 (Ac-APE1/ Ref-1)	RAGE-mediated apoptosis induction through unknown mechanisms	Breast cancer	[148]

For compounds that inhibit NF- κ B, further studies may be warranted to examine the pathways involved in apoptosis (or inhibition of cell proliferation) induction through nuclear factor κ B in cancer cells

cancer cells either directly or indirectly. Table 1 summarizes different RAGE inhibitors that can be further explored for a possible intervention in anticancer therapy.

Conclusion

The accumulation of AGE and RAGE orchestrate diverse set of signal transduction pathways for progression of oxidative stress to manifest pathological key events. Nevertheless, dysregulated activation of various set of genes and protein directly or indirectly influences cellular homeostasis via accelerated generation of free radicals and ROS. An accumulated ROS favors production of AGEs and activation of nuclear transcription factor NF-kB, with increases expression and release of pro-inflammatory cytokines resulting in the cellular damage, which triggers the release of intracellular component to the rapid activation of an array of signaling cascade culminating tumor microenvironment for cell survival. The tumor microenvironment is mainly comprised of cancer cells surrounded by inflammatory milieu with hyperglycemic and hypoxic environment which concomitantly enhances the formation of AGEs. The association of hyperglycemia and AGEs formation during cancer progression requires extensive research to decode the complex mechanism of cell proliferation of cancer cells. Beside, that the upregulation of RAGE expression is well documented in various pathological conditions. The binding of AGEs with their receptor RAGE augments oxidative stress and inflammation that collectively promotes tumorigenesis. Moreover, the programmed cell death pathway is dysregulated during cancer progression. Based on the available reports discussed above, it is inclined to believe that AGEs-RAGE signaling promotes survival pathways in cancer cells by negative feedback regulation of apoptosis and positive regulation of pro-survival mechanism such as autophagy and necroptosis mediated release of DAMPs. Moreover AGEs-RAGE signaling-mediated pro-survival autophagy limits apoptosis in cancer cells. However, in normal cells, the activation of AGEs-RAGE signaling induces cell death to balance tissue homeostasis. Dysregulated activation of AGE-RAGE signaling may lead to pathological consequences. The paradigm of AGEs-RAGE signaling-mediated diversion from cell death (apoptosis) to cell survival (pro-survival autophagy) requires extensive research. An inhibition of AGE-RAGE signaling may be beneficial to counteract progression of inflammatory cascades responsible for various pathological conditions. Further deciphering sequence of events involved in regulation of AGE production and identification of potential inhibitors of AGEs/RAGE may be of immense benefit for cancer prevention and therapeutics.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflicts of interest.

References

- Chen J-H, Lin X, Bu C, Zhang X (2018) Role of advanced glycation end products in mobility and considerations in possible dietary and nutritional intervention strategies. Nutr Metab 15:72
- Chhipa AS, Borse SP, Baksi R, Lalotra S, Nivsarkar M (2019) Targeting receptors of advanced glycation end products (RAGE): preventing diabetes induced cancer and diabetic complications. Pathol Res Pract 215:152643. https://doi.org/10.1016/j. prp.2019.152643
- Singh VP, Bali A, Singh N, Jaggi AS (2014) Advanced glycation end products and diabetic complications. Korean J Physiol Pharmacol 18:1–14. https://doi.org/10.4196/kjpp.2014.18.1.1
- Ramasamy R, Yan SF, Schmidt AM (2011) Receptor for AGE (RAGE): signaling mechanisms in the pathogenesis of diabetes and its complications. Ann N Y Acad Sci 1243:88–102. https ://doi.org/10.1111/j.1749-6632.2011.06320.x
- Senatus LM, Schmidt AM (2017) The AGE-RAGE axis: implications for age-associated arterial diseases. Front Gent 8:187
- Soman S, Raju R, Sandhya VK, Advani J, Khan AA, Harsha HC, Prasad TSK, Sudhakaran PR, Pandey A, Adishesha PK (2013) A multicellular signal transduction network of AGE/ RAGE signaling. J Cell Commun Signal 7:19–23. https://doi. org/10.1007/s12079-012-0181-3
- Hudson BI, Lippman ME (2018) Targeting RAGE signaling in inflammatory disease. Annu Rev Med 69:349–364. https://doi. org/10.1146/annurev-med-041316-085215
- Clavel J (2007) Progress in the epidemiological understanding of gene-environment interactions in major diseases: cancer. CR Biol 330:306–317. https://doi.org/10.1016/j.crvi.2007.02.012
- 9. Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. Cell 144:646–674
- Landskron G, De la Fuente M, Thuwajit P, Thuwajit C, Hermoso MA (2014) Chronic inflammation and cytokines in the tumor microenvironment. J Immunol Res 2014:149185
- Blaylock RL (2015) Cancer microenvironment, inflammation and cancer stem cells: a hypothesis for a paradigm change and new targets in cancer control. Surg Neurol Int 6:92
- Liou G-Y, Storz P (2010) Reactive oxygen species in cancer. Free Radical Res 44:479–496
- Grivennikov SI, Greten FR, Karin M (2010) Immunity, inflammation, and cancer. Cell 140:883–899
- Colotta F, Allavena P, Sica A, Garlanda C, Mantovani A (2009) Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. Carcinogenesis 30:1073–1081
- Multhoff G, Molls M, Radons J (2012) Chronic inflammation in cancer development. Front Immunol 2:98–98. https://doi. org/10.3389/fimmu.2011.00098
- Ryu TY, Park J, Scherer PE (2014) Hyperglycemia as a risk factor for cancer progression. Diab Metab J 38:330–336

- 17. Kang R, Tang D, Schapiro NE, Livesey KM, Farkas A, Loughran P, Bierhaus A, Lotze MT, Zeh HJ (2010) The receptor for advanced glycation end products (RAGE) sustains autophagy and limits apoptosis, promoting pancreatic tumor cell survival. Cell Death Differ 17:666–676
- El-Far AH, Sroga G, Jaouni SKA, Mousa SA (2020) Role and mechanisms of RAGE-ligand complexes and RAGE-inhibitors in cancer progression. Int J Mol Sci. https://doi.org/10.3390/ ijms21103613
- Riehl A, Németh J, Angel P, Hess J (2009) The receptor RAGE: bridging inflammation and cancer. Cell Commun Signal 7:12–12. https://doi.org/10.1186/1478-811X-7-12
- Nasser MW, Ahirwar DK, Ganju RK (2016) RAGE: a novel target for breast cancer growth and metastasis. Oncoscience 3:52–53. https://doi.org/10.18632/oncoscience.294
- 21. Rojas A, Figueroa H, Morales E (2010) Fueling inflammation at tumor microenvironment: the role of multiligand/RAGE axis. Carcinogenesis 31:334–341
- Khan MI, Rath S, Adhami VM and Mukhar H (2018) Hypoxia driven glycation: mechanisms and therapeutic opportunities. Semin Cancer Biol 49:75–82. https://doi.org/10.1016/j.semca ncer.2017.05.008
- 23. Kang R, Hou W, Zhang Q, Chen R, Lee Y, Bartlett D, Lotze M, Tang D, Zeh H (2014) RAGE is essential for oncogenic KRAS-mediated hypoxic signaling in pancreatic cancer. Cell Death Dis 5:e1480
- 24. EI-Far A (2016) The role of receptors for advanced glycation end product in pancreatic carcinogenesis. Pancreat Disord Ther 6:166
- 25. Chen M-C, Chen K-C, Chang G-C, Lin H, Wu C-C, Kao W-H, Teng C-LJ, Hsu S-L, Yang T-Y (2020) RAGE acts as an oncogenic role and promotes the metastasis of human lung cancer. Cell Death Dise 11:1–13
- Matou-Nasri S, Sharaf H, Wang Q, Almobadel N, Rabhan Z, Al-Eidi H, Yahya WB, Trivilegio T, Ali R, Al-Shanti N (2017) Biological impact of advanced glycation endproducts on estrogen receptor-positive MCF-7 breast cancer cells. Biochim Biophys Acta BBA 1863:2808–2820
- Xu X, Abuduhadeer X, Zhang W, Li T, Gao H, Wang Y (2013) Knockdown of RAGE inhibits growth and invasion of gastric cancer cells. Eur J Histochem 57:e36
- Swami P, Radhakrishnan P, Crawford A, Patil P, Shin S, Caffrey T, Grunkemeyer J, O'connell K, Hollingsworth M, Leclerc E (2019) Combination of RAGE inhibitors and gemcitabine impedes tumor growth by reducing autophagy and facilitating apoptosis in pancreatic cancer. FASEP J 33:674.19
- Kwak T, Drews-Elger K, Ergonul A, Miller P, Braley A, Hwang G, Zhao D, Besser A, Yamamoto Y, Yamamoto H (2017) Targeting of RAGE-ligand signaling impairs breast cancer cell invasion and metastasis. Oncogene 36:1559–1572
- 30. Ishiguro H, Nakaigawa N, Miyoshi Y, Fujinami K, Kubota Y, Uemura H (2005) Receptor for advanced glycation end products (RAGE) and its ligand, amphoterin are overexpressed and associated with prostate cancer development. Prostate 64:92–100
- Redza-Dutordoir M, Averill-Bates DA (2016) Activation of apoptosis signalling pathways by reactive oxygen species. Biochim Biophys Acta BBA 1863:2977–2992
- Sies H, Jones DP (2020) Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. Nat Rev Mol Cell Biol 21:1–21. https://doi.org/10.1038/s41580-020-0230-3
- Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, Gargiulo G, Testa G, Cacciatore F, Bonaduce D (2018) Oxidative stress, aging, and diseases. Clin Interv Aging 13:757
- Palanissami G, Paul SF (2018) RAGE and its ligands: molecular interplay between glycation, inflammation, and hallmarks of cancer—a review. Horm Cancer 9:295–325

- Tan AL, Forbes JM, Cooper ME (2007) AGE, RAGE, and ROS in diabetic nephropathy. Semin Nephrol 27:130–143. https://doi. org/10.1016/j.semnephrol.2007.01.006
- Nowotny K, Jung T, Höhn A, Weber D, Grune T (2015) Advanced glycation end products and oxidative stress in type 2 diabetes mellitus. Biomolecules 5:194–222
- Warboys CM, Toh H-B, Fraser PA (2009) Role of NADPH oxidase in retinal microvascular permeability increase by RAGE activation. Invest Ophthalmol Vis Sci 50:1319–1328
- Sun Y, Peng Z (2009) Programmed cell death and cancer. Postgrad Med J 85:134–140
- 39. Gewies A (2003) Introduction to apoptosis. Apo Rev 3:1-26
- Denton D, Kumar S (2019) Autophagy-dependent cell death. Cell Death Differ 26:605–616
- Choi ME, Price DR, Ryter SW, Choi AM (2019) Necroptosis: a crucial pathogenic mediator of human disease. JCI Insight 4:e128834
- 42. Jo Y, Choi N, Kim K, Koo H-J, Choi J, Kim HN (2018) Chemoresistance of cancer cells: requirements of tumor microenvironment-mimicking in vitro models in anti-cancer drug development. Theranostics 8:5259
- Kim S, Kwon J (2013) COMP-Ang1 inhibits apoptosis as well as improves the attenuated osteogenic differentiation of mesenchymal stem cells induced by advanced glycation end products. Biochim Biophys Acta 1830:4928–4934. https://doi.org/10.1016/j. bbagen.2013.06.035
- Barlovic DP, Thomas MC, Jandeleit-Dahm K (2010) Cardiovascular disease: what's all the AGE/RAGE about? Cardiovasc Hematol Disord Drug Targets 10:7–15
- 45. Zhan Y, Sun HL, Chen H, Zhang H, Sun J, Zhang Z, Cai DH (2012) Glucagon-like peptide-1 (GLP-1) protects vascular endothelial cells against advanced glycation end products (AGEs)-induced apoptosis. Med Sci Monit 18:286–291. https:// doi.org/10.12659/msm.883207
- 46. Kim J, Kim KM, Kim CS, Sohn E, Lee YM, Jo K, Kim JS (2012) Puerarin inhibits the retinal pericyte apoptosis induced by advanced glycation end products in vitro and in vivo by inhibiting NADPH oxidase-related oxidative stress. Free Radic Biol Med 53:357–365. https://doi.org/10.1016/j.freeradbio med.2012.04.030
- 47. Gao Y, Wake H, Morioka Y, Liu K, Teshigawara K, Shibuya M, Zhou J, Mori S, Takahashi H, Nishibori M (2017) Phagocytosis of advanced glycation end products (AGEs) in macrophages induces cell apoptosis. Oxid Med Cell Longev 2017:8419035
- Strasser A, O'Connor L, Dixit VM (2000) Apoptosis signaling. Annu Rev Biochem 69:217–245. https://doi.org/10.1146/annur ev.biochem.69.1.217
- Boulanger E, Wautier MP, Wautier JL, Boval B, Panis Y, Wernert N, Danze PM, Dequiedt P (2002) AGEs bind to mesothelial cells via RAGE and stimulate VCAM-1 expression. Kidney Int 61:148–156. https://doi.org/10.1046/j.1523-1755.2002.00115.x
- Xie J, Mendez JD, Mendez-Valenzuela V, Aguilar-Hernandez MM (2013) Cellular signalling of the receptor for advanced glycation end products (RAGE). Cell Signal 25:2185–2197. https ://doi.org/10.1016/j.cellsig.2013.06.013
- Tsoporis JN, Izhar S, Leong-Poi H, Desjardins JF, Huttunen HJ, Parker TG (2010) S100B interaction with the receptor for advanced glycation end products (RAGE): a novel receptor-mediated mechanism for myocyte apoptosis postinfarction. Circ Res 106:93–101. https://doi.org/10.1161/CIRCRESAHA.109.19583 4
- 52. Wang P, Xing Y, Chen C, Chen Z, Qian Z (2016) Advanced glycation end-product (AGE) induces apoptosis in human retinal ARPE-19 cells via promoting mitochondrial dysfunction and activating the Fas-FasL signaling. Biosci Biotechnol Biochem 80:250–256. https://doi.org/10.1080/09168451.2015.1095065

- 53. Taniguchi N, Kawahara K, Yone K, Hashiguchi T, Yamakuchi M, Goto M, Inoue K, Yamada S, Ijiri K, Matsunaga S, Nakajima T, Komiya S, Maruyama I (2003) High mobility group box chromosomal protein 1 plays a role in the pathogenesis of rheumatoid arthritis as a novel cytokine. Arthritis Rheum 48:971–981. https://doi.org/10.1002/art.10859
- Fiuza C, Bustin M, Talwar S, Tropea M, Gerstenberger E, Shelhamer JH, Suffredini AF (2003) Inflammation-promoting activity of HMGB1 on human microvascular endothelial cells. Blood 101:2652–2660. https://doi.org/10.1182/blood -2002-05-1300
- Lecomte M, Denis U, Ruggiero D, Lagarde M, Wiernsperger N (2004) Involvement of caspase-10 in advanced glycation endproduct-induced apoptosis of bovine retinal pericytes in culture. Biochim Biophys Acta 1689:202–211. https://doi.org/10.1016/j. bbadis.2004.03.010
- Denis U, Lecomte M, Paget C, Ruggiero D, Wiernsperger N, Lagarde M (2002) Advanced glycation end-products induce apoptosis of bovine retinal pericytes in culture: involvement of diacylglycerol/ceramide production and oxidative stress induction. Free Radic Biol Med 33:236–247. https://doi.org/10.1016/ s0891-5849(02)00879-1
- 57. Pan Y, Liang H, Liu H, Li D, Chen X, Li L, Zhang CY, Zen K (2014) Platelet-secreted microRNA-223 promotes endothelial cell apoptosis induced by advanced glycation end products via targeting the insulin-like growth factor 1 receptor. J Immunol 192:437–446. https://doi.org/10.4049/jimmunol.1301790
- 58. Sun C, Liang C, Ren Y, Zhen Y, He Z, Wang H, Tan H, Pan X, Wu Z (2009) Advanced glycation end products depress function of endothelial progenitor cells via p38 and ERK 1/2 mitogenactivated protein kinase pathways. Basic Res Cardiol 104:42–49. https://doi.org/10.1007/s00395-008-0738-8
- Ott C, Jacobs K, Haucke E, Navarrete Santos A, Grune T, Simm A (2014) Role of advanced glycation end products in cellular signaling. Redox Biol 2:411–429. https://doi.org/10.1016/j.redox .2013.12.016
- Alikhani M, Maclellan CM, Raptis M, Vora S, Trackman PC, Graves DT (2007) Advanced glycation end products induce apoptosis in fibroblasts through activation of ROS, MAP kinases, and the FOXO1 transcription factor. Am J Physiol Cell Physiol 292:C850–C856. https://doi.org/10.1152/ajpcell.00356.2006
- Kumar S, Boehm J, Lee JC (2003) p38 MAP kinases: key signalling molecules as therapeutic targets for inflammatory diseases. Nat Rev Drug Discov 2:717–726. https://doi.org/10.1038/nrd11 77
- Liu J, Lin A (2005) Role of JNK activation in apoptosis: a double-edged sword. Cell Res 15:36–42. https://doi.org/10.1038/ sj.cr.7290262
- Fang D, Hawke D, Zheng Y, Xia Y, Meisenhelder J, Nika H, Mills GB, Kobayashi R, Hunter T, Lu Z (2007) Phosphorylation of beta-catenin by AKT promotes beta-catenin transcriptional activity. J Biol Chem 282:11221–11229. https://doi.org/10.1074/ jbc.M611871200
- 64. Huang HL, Hsieh MJ, Chien MH, Chen HY, Yang SF, Hsiao PC (2014) Glabridin mediate caspases activation and induces apoptosis through JNK1/2 and p38 MAPK pathway in human promyelocytic leukemia cells. PLoS ONE 9:e98943. https://doi. org/10.1371/journal.pone.0098943
- 65. Alikhani M, Alikhani Z, Boyd C, MacLellan CM, Raptis M, Liu R, Pischon N, Trackman PC, Gerstenfeld L, Graves DT (2007) Advanced glycation end products stimulate osteoblast apoptosis via the MAP kinase and cytosolic apoptotic pathways. Bone 40:345–353. https://doi.org/10.1016/j.bone.2006.09.011
- 66. Shi L, Yu X, Yang H, Wu X (2013) Advanced glycation end products induce human corneal epithelial cells apoptosis through generation of reactive oxygen species and activation of JNK

and p38 MAPK pathways. PLoS ONE 8:e66781. https://doi.org/10.1371/journal.pone.0066781

- 67. Liu BF, Miyata S, Hirota Y, Higo S, Miyazaki H, Fukunaga M, Hamada Y, Ueyama S, Muramoto O, Uriuhara A, Kasuga M (2003) Methylglyoxal induces apoptosis through activation of p38 mitogen-activated protein kinase in rat mesangial cells. Kidney Int 63:947–957. https://doi.org/10.104 6/j.1523-1755.2003.00829.x
- Sekido H, Suzuki T, Jomori T, Takeuchi M, Yabe-Nishimura C, Yagihashi S (2004) Reduced cell replication and induction of apoptosis by advanced glycation end products in rat Schwann cells. Biochem Biophys Res Commun 320:241–248. https://doi. org/10.1016/j.bbrc.2004.05.159
- Alikhani Z, Alikhani M, Boyd CM, Nagao K, Trackman PC, Graves DT (2005) Advanced glycation end products enhance expression of pro-apoptotic genes and stimulate fibroblast apoptosis through cytoplasmic and mitochondrial pathways. J Biol Chem 280:12087–12095. https://doi.org/10.1074/jbc.M4063 13200
- Kowluru RA (2005) Effect of advanced glycation end products on accelerated apoptosis of retinal capillary cells under in vitro conditions. Life Sci 76:1051–1060. https://doi.org/10.1016/j. lfs.2004.10.017
- Min C, Kang E, Yu SH, Shinn SH, Kim YS (1999) Advanced glycation end products induce apoptosis and procoagulant activity in cultured human umbilical vein endothelial cells. Diab Res Clin Pract 46:197–202
- Riuzzi F, Sorci G, Donato R (2006) The amphoterin (HMGB1)/ receptor for advanced glycation end products (RAGE) pair modulates myoblast proliferation, apoptosis, adhesiveness, migration, and invasiveness. Functional inactivation of RAGE in L6 myoblasts results in tumor formation in vivo. J Biol Chem 281:8242– 8253. https://doi.org/10.1074/jbc.M509436200
- Krysko O, Love Aaes T, Bachert C, Vandenabeele P, Krysko DV (2013) Many faces of DAMPs in cancer therapy. Cell Death Dis 4:e631. https://doi.org/10.1038/cddis.2013.156
- Tang D, Kang R, Cheh CW, Livesey KM, Liang X, Schapiro NE, Benschop R, Sparvero LJ, Amoscato AA, Tracey KJ, Zeh HJ, Lotze MT (2010) HMGB1 release and redox regulates autophagy and apoptosis in cancer cells. Oncogene 29:5299–5310. https:// doi.org/10.1038/onc.2010.261
- Zhang X, Chen Y, Cai G, Li X, Wang D (2017) Carnosic acid induces apoptosis of hepatocellular carcinoma cells via ROSmediated mitochondrial pathway. Chem Biol Interact 277:91– 100. https://doi.org/10.1016/j.cbi.2017.09.005
- 76. Tse AK, Cao HH, Cheng CY, Kwan HY, Yu H, Fong WF, Yu ZL (2014) Indomethacin sensitizes TRAIL-resistant melanoma cells to TRAIL-induced apoptosis through ROS-mediated upregulation of death receptor 5 and downregulation of survivin. J Invest Dermatol 134:1397–1407. https://doi.org/10.1038/jid.2013.471
- 77. Lo MC, Chen MH, Lee WS, Lu CI, Chang CR, Kao SH, Lee HM (2015) Nepsilon-(carboxymethyl) lysine-induced mitochondrial fission and mitophagy cause decreased insulin secretion from beta-cells. Am J Physiol Endocrinol Metab 309:E829–E839. https://doi.org/10.1152/ajpendo.00151.2015
- 78. Yu Y, Wang L, Delguste F, Durand A, Guilbaud A, Rousselin C, Schmidt AM, Tessier F, Boulanger E, Neviere R (2017) Advanced glycation end products receptor RAGE controls myocardial dysfunction and oxidative stress in high-fat fed mice by sustaining mitochondrial dynamics and autophagy-lysosome pathway. Free Radic Biol Med 112:397–410. https://doi. org/10.1016/j.freeradbiomed.2017.08.012
- 79. Mao YX, Cai WJ, Sun XY, Dai PP, Li XM, Wang Q, Huang XL, He B, Wang PP, Wu G, Ma JF, Huang SB (2018) RAGE-dependent mitochondria pathway: a novel target of silibinin against apoptosis of osteoblastic cells induced by advanced glycation end

products. Cell Death Dis 9:674. https://doi.org/10.1038/s4141 9-018-0718-3

- Kang R, Tang D, Schapiro NE, Loux T, Livesey KM, Billiar TR, Wang H, Van Houten B, Lotze MT, Zeh HJ (2014) The HMGB1/ RAGE inflammatory pathway promotes pancreatic tumor growth by regulating mitochondrial bioenergetics. Oncogene 33:567– 577. https://doi.org/10.1038/onc.2012.631
- Xu L, Fan Q, Wang X, Zhao X, Wang L (2016) Inhibition of autophagy increased AGE/ROS-mediated apoptosis in mesangial cells. Cell Death Dis 7:e2445. https://doi.org/10.1038/cddis .2016.322
- Yamagishi S, Inagaki Y, Okamoto T, Amano S, Koga K, Takeuchi M, Makita Z (2002) Advanced glycation end productinduced apoptosis and overexpression of vascular endothelial growth factor and monocyte chemoattractant protein-1 in humancultured mesangial cells. J Biol Chem 277:20309–20315. https ://doi.org/10.1074/jbc.M202634200
- Hung LF, Huang KY, Yang DH, Chang DM, Lai JH, Ho LJ (2010) Advanced glycation end products induce T cell apoptosis: involvement of oxidative stress, caspase and the mitochondrial pathway. Mech Ageing Dev 131:682–691. https://doi. org/10.1016/j.mad.2010.09.005
- 84. Sparvero LJ, Asafu-Adjei D, Kang R, Tang D, Amin N, Im J, Rutledge R, Lin B, Amoscato AA, Zeh HJ (2009) RAGE (receptor for advanced glycation endproducts), RAGE ligands, and their role in cancer and inflammation. J Transl Med 7:17
- Dhumale SS, Waghela BN, Pathak C (2015) Quercetin protects necrotic insult and promotes apoptosis by attenuating the expression of RAGE and its ligand HMGB1 in human breast adenocarcinoma cells. IUBMB Life 67:361–373
- Kang R, Tang D, Lotze MT, Zeh I, Herbert J (2011) RAGE regulates autophagy and apoptosis following oxidative injury. Autophagy 7:442–444
- Zhang Q-Y, Wu L-Q, Zhang T, Han Y-F, Lin X (2015) Autophagy-mediated HMGB1 release promotes gastric cancer cell survival via RAGE activation of extracellular signal-regulated kinases 1/2. Oncol Rep 33:1630–1638
- Cheng P, Dai W, Wang F, Lu J, Shen M, Chen K, Li J, Zhang Y, Wang C, Yang J (2014) Ethyl pyruvate inhibits proliferation and induces apoptosis of hepatocellular carcinoma via regulation of the HMGB1–RAGE and AKT pathways. Biochem Biophys Res Commun 443:1162–1168
- Elangovan I, Thirugnanam S, Chen A, Zheng G, Bosland MC, Kajdacsy-Balla A, Gnanasekar M (2012) Targeting receptor for advanced glycation end products (RAGE) expression induces apoptosis and inhibits prostate tumor growth. Biochem Biophys Res Commun 417:1133–1138
- Lan C-Y, Chen S-Y, Kuo C-W, Lu C-C, Yen G-C (2019) Quercetin facilitates cell death and chemosensitivity through RAGE/ PI3K/AKT/mTOR axis in human pancreatic cancer cells. J Food Drug Anal 27:887–896
- Kang R, Tang D, Loze MT, Zeh I, Herbert J (2011) Apoptosis to autophagy switch triggered by the MHC class III-encoded receptor for advanced glycation endproducts (RAGE). Autophagy 7:91–93
- Recouvreux MV, Commisso C (2017) Macropinocytosis: a metabolic adaptation to nutrient stress in cancer. Front Endocrinol (Lausanne) 8:261. https://doi.org/10.3389/fendo.2017.00261
- Schmukler E, Kloog Y, Pinkas-Kramarski R (2014) Ras and autophagy in cancer development and therapy. Oncotarget 5:577–586. https://doi.org/10.18632/oncotarget.1775
- Yun CW, Lee SH (2018) The roles of autophagy in cancer. Int J Mol Sci. https://doi.org/10.3390/ijms19113466
- Daskalaki I, Gkikas I, Tavernarakis N (2018) Hypoxia and selective autophagy in cancer development and therapy. Front Cell Dev Biol 6:104. https://doi.org/10.3389/fcell.2018.00104

- Levy JM, Thompson JC, Griesinger AM, Amani V, Donson AM, Birks DK, Morgan MJ, Mirsky DM, Handler MH, Foreman NK, Thorburn A (2014) Autophagy inhibition improves chemosensitivity in BRAF(V600E) brain tumors. Cancer Discov 4:773–780. https://doi.org/10.1158/2159-8290.CD-14-0049
- Cho SW, Na W, Choi M, Kang SJ, Lee S-G, Choi CY (2017) Autophagy inhibits cell death induced by the anti-cancer drug morusin. Am J Cancer Res 7:518–530
- Lane JD, Carroll B, Hewitt G, Korolchuk VI (2013) Autophagy and ageing: implications for age-related neurodegenerative diseases. Essays Biochem 55:119–131
- 99. Hu P, Lai D, Lu P, Gao J, He H (2012) ERK and Akt signaling pathways are involved in advanced glycation end productinduced autophagy in rat vascular smooth muscle cells. Int J Mol Med 29:613–618
- 100. Kang R, Tang D, Livesey KM, Schapiro NE, Lotze MT, Zeh HJ 3rd (2011) The receptor for advanced glycation end-products (RAGE) protects pancreatic tumor cells against oxidative injury. Antioxid Redox Signal 15:2175–2184. https://doi.org/10.1089/ ars.2010.3378
- Piperi C, Adamopoulos C, Papavassiliou AG (2017) Potential of glycative stress targeting for cancer prevention. Cancer Lett 390:153–159. https://doi.org/10.1016/j.canlet.2017.01.020
- 102. Verma N, Manna SK (2016) Advanced glycation end products (AGE) potently induce autophagy through activation of RAF protein kinase and nuclear factor kappaB (NF-kappaB). J Biol Chem 291:1481–1491. https://doi.org/10.1074/jbc.M115.66757 6
- 103. He M, Siow RC, Sugden D, Gao L, Cheng X, Mann GE (2011) Induction of HO-1 and redox signaling in endothelial cells by advanced glycation end products: a role for Nrf2 in vascular protection in diabetes. Nutr Metab Cardiovasc Dis 21:277–285
- 104. Ko SY, Ko HA, Shieh TM, Chi TC, Chen HI, Chen YT, Yu YH, Yang SH, Chang SS (2017) Advanced glycation end products influence oral cancer cell survival via Bcl-xl and Nrf-2 regulation in vitro. Oncol Lett 13:3328–3334
- 105. Gao W, Zhou Z, Liang B, Huang Y, Yang Z, Chen Y, Zhang L, Yan C, Wang J, Lu L, Wen Z, Xian S, Wang L (2018) Inhibiting receptor of advanced glycation end products attenuates pressure overload-induced cardiac dysfunction by preventing excessive autophagy. Front Physiol 9:1333. https://doi.org/10.3389/fphys .2018.01333
- 106. Hu P, Zhou H, Lu M, Dou L, Bo G, Wu J, Huang S (2015) Autophagy plays a protective role in advanced glycation end product-induced apoptosis in cardiomyocytes. Cell Physiol Biochem 37:697–706. https://doi.org/10.1159/000430388
- 107. Hou X, Hu Z, Xu H, Xu J, Zhang S, Zhong Y, He X, Wang N (2014) Advanced glycation endproducts trigger autophagy in cadiomyocyte via RAGE/PI3K/AKT/mTOR pathway. Cardiovasc Diabetol 13:78. https://doi.org/10.1186/1475-2840-13-78
- 108. Li J, Wu PW, Zhou Y, Dai B, Zhang PF, Zhang YH, Liu Y, Shi XL (2018) Rage induces hepatocellular carcinoma proliferation and sorafenib resistance by modulating autophagy. Cell Death Dis 9:225. https://doi.org/10.1038/s41419-018-0329-z
- 109. Gong Y, Fan Z, Luo G, Yang C, Huang Q, Fan K, Cheng H, Jin K, Ni Q, Yu X (2019) The role of necroptosis in cancer biology and therapy. Mol Cancer 18:100
- LaRocca TJ, Sosunov SA, Shakerley NL, Ten VS, Ratner AJ (2016) Hyperglycemic conditions prime cells for RIP1-dependent necroptosis. J Biol Chem 291:13753–13761
- 111. Su Z, Yang Z, Xu Y, Chen Y, Yu Q (2015) Apoptosis, autophagy, necroptosis, and cancer metastasis. Mol Cancer 14:48
- 112. Moreno-Gonzalez G, Vandenabeele P, Krysko DV (2016) Necroptosis: a novel cell death modality and its potential relevance for critical care medicine. Am J Respir Crit Care Med 194:415–428

- 113. Nogueira-Machado JA, Volpe CMDO, Veloso CA, Chaves MMJ (2011) HMGB1, TLR and RAGE: a functional tripod that leads to diabetic inflammation. Expert Opin Ther Targets 15:1023–1035
- 114. Wu L, Yang L (2018) The function and mechanism of HMGB1 in lung cancer and its potential therapeutic implications. Oncol Lett 15:6799–6805
- 115. Li X, Jiang S, Tapping RI (2010) Toll-like receptor signaling in cell proliferation and survival. Cytokine 49:1–9
- Nogueira-Machado JA, Volpe CMDO, Veloso CA, Chaves MM (2011) HMGB1, TLR and RAGE: a functional tripod that leads to diabetic inflammation. Exp Opin Ther Targets 15:1023–1035
- 117. Akira S, Takeda K (2004) Toll-like receptor signalling. Nat Rev Immunol 4:499–511
- 118. Zhang C, Dong H, Chen F, Wang Y, Ma J, Wang G (2019) The HMGB1-RAGE/TLR-TNF-α signaling pathway may contribute to kidney injury induced by hypoxia. Exp Ther Med 17:17–26
- 119. Alcalá S, Sainz B (2019) The dark side of radiotherapy-induced cell death in cancer. EBioMedicine 40:7–8
- O'Neill LA, Bowie AG (2007) The family of five: TIR-domaincontaining adaptors in Toll-like receptor signalling. Nat Rev Immunol 7:353–364
- 121. Sims GP, Rowe DC, Rietdijk ST, Herbst R, Coyle AJ (2009) HMGB1 and RAGE in inflammation and cancer. Annu Rev Immunol 28:367–388
- Tang D, Kang R, Zeh HJ III, Lotze MT (2010) High-mobility group box 1 and cancer. Biochem Biophys Acta 1799:131–140
- 123. Zhang T, Guan X-W, Gribben JG, Liu F-T, Jia L (2019) Blockade of HMGB1 signaling pathway by ethyl pyruvate inhibits tumor growth in diffuse large B-cell lymphoma. Cell Death Dis 10:1–15
- Bartel DP (2004) MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 116:281–297
- 125. Ambros V (2004) The functions of animal microRNAs. Nature 431:350–355
- Alvarez-Garcia I, Miska EA (2005) MicroRNA functions in animal development and human disease. Development 132:4653–4662
- Zhang B, Pan X, Cobb GP, Anderson TA (2007) microRNAs as oncogenes and tumor suppressors. Dev Biol 302:1–12
- Bracken CP, Scott HS, Goodall GJ (2016) A network-biology perspective of microRNA function and dysfunction in cancer. Nat Rev Genet 17:719–732
- Choudhury Y, Tay FC, Lam DH, Sandanaraj E, Tang C, Ang B-T, Wang S (2012) Attenuated adenosine-to-inosine editing of microRNA-376a* promotes invasiveness of glioblastoma cells. J Clin Investig 122:4059–4076
- Lin S, Gregory RI (2015) MicroRNA biogenesis pathways in cancer. Nat Rev Cancer 15:321–333
- 131. Dvinge H, Git A, Gräf S, Salmon-Divon M, Curtis C, Sottoriva A, Zhao Y, Hirst M, Armisen J, Miska EA (2013) The shaping and functional consequences of the microRNA landscape in breast cancer. Nature 497:378–382
- 132. Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, Sweet-Cordero A, Ebert BL, Mak RH, Ferrando AA (2005) MicroRNA expression profiles classify human cancers. Nature 435:834–838
- 133. Mercado-Pimentel ME, Onyeagucha BC, Li Q, Pimentel AC, Jandova J, Nelson MA (2015) The S100P/RAGE signaling pathway regulates expression of microRNA-21 in colon cancer cells. FEBS Lett 589:2388–2393
- 134. Faltejskova P, Besse A, Sevcikova S, Kubiczkova L, Svoboda M, Smarda J, Kiss I, Vyzula R, Slaby O (2012) Clinical correlations of miR-21 expression in colorectal cancer patients and effects of its inhibition on DLD1 colon cancer cells. Int J Colorectal Dis 27:1401–1408

- 135. Liu X, Cheng Y, Zhang S, Lin Y, Yang J, Zhang C (2009) A necessary role of miR-221 and miR-222 in vascular smooth muscle cell proliferation and neointimal hyperplasia. Circ Res 104:476–487
- 136. Mardente S, Mari E, Consorti F, Di Gioia C, Negri R, Etna M, Zicari A, Antonaci A (2012) HMGB1 induces the overexpression of miR-222 and miR-221 and increases growth and motility in papillary thyroid cancer cells. Oncol Rep 28:2285–2289
- 137. Mardente S, Mari E, Massimi I, Fico F, Faggioni A, Pulcinelli F, Antonaci A, Zicari A (2015) HMGB1-induced cross talk between PTEN and miRs 221/222 in thyroid cancer. Biomed Res Int 2015:512027
- Li L-M, Hou D-X, Guo Y-L, Yang J-W, Liu Y, Zhang C-Y, Zen K (2011) Role of microRNA-214–targeting phosphatase and tensin homolog in advanced glycation end product-induced apoptosis delay in monocytes. J Immunol 186:2552–2560
- 139. Qin Y, Ye J, Wang P, Gao L, Wang S, Shen H (2016) miR-223 contributes to the AGE-promoted apoptosis via down-regulating insulin-like growth factor 1 receptor in osteoblasts. Biosci Rep 36:e00314
- 140. Wang L, Kang F-B, Wang J, Yang C, He D-W (2019) Downregulation of miR-205 contributes to epithelial–mesenchymal transition and invasion in triple-negative breast cancer by targeting HMGB1–RAGE signaling pathway. Anticancer Drugs 30:225
- 141. El-Far AHAM, Munesue S, Harashima A, Sato A, Shindo M, Nakajima S, Inada M, Tanaka M, Takeuchi A, Tsuchiya H (2018) In vitro anticancer effects of a RAGE inhibitor discovered using a structure-based drug design system. Oncol Lett 15:4627–4634
- 142. Jimenez R, Lopez-Sepulveda R, Romero M, Toral M, Cogolludo A, Perez-Vizcaino F, Duarte J (2015) Quercetin and its metabolites inhibit the membrane NADPH oxidase activity in vascular smooth muscle cells from normotensive and spontaneously hypertensive rats. Food Funct 6:409–414
- 143. Hahm E-R, Moura MB, Kelley EE, Van Houten B, Shiva S, Singh SV (2011) Withaferin A-induced apoptosis in human breast cancer cells is mediated by reactive oxygen species. PLoS ONE 6:e23354
- 144. Park JW, Min K-J, Kim DE, Kwon TK (2015) Withaferin A induces apoptosis through the generation of thiol oxidation in human head and neck cancer cells. Int J Mol Med 35:247–252
- 145. Arumugam T, Ramachandran V, Gomez SB, Schmidt AM, Logsdon CD (2012) S100P-derived RAGE antagonistic peptide reduces tumor growth and metastasis. Clin Cancer Res 18:4356–4364
- 146. Song T-Y, Yang N-C, Chen C-L, Thi TLV (2017) Protective effects and possible mechanisms of ergothioneine and hispidin against methylglyoxal-induced injuries in rat pheochromocytoma cells. Oxid Med Cell Longev. https://doi.org/10.1155/2017/48243 71
- 147. Pellegrini L, Xue J, Larson D, Pastorino S, Jube S, Forest KH, Saad-Jube ZS, Napolitano A, Pagano I, Negi VS (2017) HMGB1 targeting by ethyl pyruvate suppresses malignant phenotype of human mesothelioma. Oncotarget 8:22649
- 148. Lee YR, Kim KM, Jeon BH, Choi S (2015) Extracellularly secreted APE1/Ref-1 triggers apoptosis in triple-negative breast cancer cells via RAGE binding, which is mediated through acetylation. Oncotarget 6:23383

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